



JURNAL TEKNOLOGI LABORATORIUM

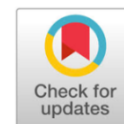
Journal Homepage: www.teknolabjournal.com
 ISSN 2580-0191(Online) | ISSN 2338 – 5634(Print)



Original Research



Decreasing α -synuclein Aggregation by Ethanol Extract of Keluwih (*Artocarpus camansi*) Leaves on Rotenone-Induced Adult Zebrafish as Parkinson's Diseases Model



Marisca Evalina Gondokesumo ¹, Faisal Akhmal Muslikh ²,
 Ni Putu Diah Nopitasari ¹, Putu Dea Angelita Putri ¹

¹ Faculty of Pharmacy, University of Surabaya, Surabaya, Indonesia

² Faculty of Pharmacy, Universitas Airlangga, Surabaya, Indonesia

Abstract: The prevalence of Parkinson's disease is increasing every year. This progressive disease is characterized by the loss of neurons in the substantia nigra due to the presence of α -synuclein aggregates. Keluwih leaves (*Artocarpus camansi*) are known to have activity in inhibiting acetylcholinesterase, as well as being an antioxidant and anti-inflammatory. The aim of this study was to evaluate the effect of ethanol extract of *A. camansi* leaves on the levels of α -synuclein in male and female adult zebrafish induced with rotenone. The zebrafish were induced with rotenone at a concentration of 5 μ g/L for 28 days, along with the administration of 96% ethanol extract of *A. camansi* leaves at doses of 2.5, 5, 7.5, or 10 mg/L. The media was changed every 48 hours to maintain the concentration of rotenone and extract. After 28 days, α -synuclein levels were examined using immunohistochemistry. The administration of ethanol extract of *A. camansi* leaves can reduce the average levels of α -synuclein in male and female adult zebrafish, with the optimum dose being 2.5 mg/L. Therefore, it can be concluded that the administration of ethanol extract of *A. camansi* leaves can be used as an alternative treatment for Parkinson's disease.

Keywords: α -synuclein, *Artocarpus camansi*, Parkinson disease, Zebrafis.

INTRODUCTION

The prevalence of Parkinson's disease (PD) has doubled over the last 25 years. Disability and mortality rates caused by Parkinson's disease are higher and grow faster than other neurodegenerative diseases. In 2019, Parkinson's disease morbidity rose to 5.8 million cases of disability and 329,000 cases of death.¹ This data places PD second-ranked as the most common disease, especially in people over 60 years old, then the cases are estimated to double in 2030 while increasing frequently each year.²

Parkinson's disease is characterized by the progressive loss of neurons in the substantia nigra, which is affected by decreasing dopamine production and Lewy bodies (LB) accumulation. Those conditions affect the presence of α -synuclein aggregates. The formation of LB impairs the ubiquitin-proteasome degradation process, causing mitochondrial dysfunction and then failure of adenosine triphosphate (ATP) production.³ Mitochondrial dysfunction results in abnormal regulation of calcium ions involved with the increasing calcium ion level, thus it is toxic for neurons and accumulates α -synuclein aggregates.⁴ Accordingly, mitochondrial dysfunction leads to dopaminergic damage and peripheral motor nerve degeneration in experimental animals.^{5,6}

The common animal used for the PD model is the zebrafish (*Danio rerio*) due to the uniqueness of its ventral telencephalon, which is similar to the human striatum.^{4,7} Parkinson's disease in zebrafish is induced by rotenone.⁸ Rotenone,

Corresponding author.

E-mail address: marisca@staff.ubaya.ac.id (Marisca Evalina Gondokesumo)

DOI: [10.29238/teknolabjournal.v12i2.408](https://doi.org/10.29238/teknolabjournal.v12i2.408)

Received 17 May 2023; Received in revised form 24 July 2023; Accepted 05 September 2023

© 2023 The Authors. Published by Poltekkes Kemenkes Yogyakarta, Indonesia.

This is an open-access article under the [CC BY-SA license](https://creativecommons.org/licenses/by-sa/4.0/).

one of the commonly used pesticides, is a neurotoxin that penetrates the cell membrane and causes complex I mitochondrial dysfunction and oxidative stress production,⁸ consequently dopaminergic damage and peripheral motor nerve degeneration.⁴

Traditionally, the plant *keluwih* (*Artocarpus camansi*) use in seizure treatment.⁹ *Artocarpus camansi* that belongs to the Moraceae family spreads in various countries such as Indonesia, India, Malaysia, Africa, Australia, Brazil and others. *Keluwih* leaves have known potential as antioxidants, anti-inflammatory agents, antibacterial agents, and antivirals. The phytochemicals in ethanol extract of *keluwih* leaves show several active compounds, such as flavonoids, alkaloids, tannins, triterpenoids, and phenolics (Jangtap, 2010; Solichah et al., 2021).^{10,11} Other studies also proved that several genera of *Artocarpus* including *A. camansi* contain artomunoisoxanthone, artocommunol CC, artochamin D, artochamin B, and dihydroartomunoxanthone from leaves extract, which have potential as antioxidants to fight oxidative damage.¹⁰ *Keluwih* contains compounds such as stilbenoids, arylbenzofurans, and abundant flavonoids.¹⁰ The flavonoid compounds from *keluwih* leaf extract confirmed inhibiting acetylcholinesterase (AChE) enzyme activity with anticholinergic and antioxidant effects.¹² Those properties are effective in neurodegenerative disease treatment. This study aims to evaluate the effectiveness of ethanol extract of *keluwih* leaves on α -synuclein levels in rotenone-induced adult zebrafish, both male and female, as PD patient models.

MATERIAL AND METHOD

Zebrafish

A wild type strain of black zebrafish, both male and female, were acquired from Tulungagung cultivators in East Java, Indonesia, and were identified by Airlangga University, Faculty of Fisheries and Maritime Affairs in Surabaya, East Java using identification letter number 074/ULMKILP/UA.FPK/12/2022. Late adulthood zebrafish, 9-12 months of age with 0.08 g body mass and 30.6 ± 0.95 mm length were chosen for the study to represent human brain of elderly¹³, which is the age range commonly Parkinson's disease sufferers. The zebrafish were acclimatized for seven days and were maintained in accordance to the standard procedures set by the research ethics committee of Airlangga University (No: 3.KEH.159.11.2022).

A. camansi Leaf Extraction

Keluwih leaves (*A. camansi*) were obtained from the UPT Herbal Laboratory Materia Medika Batu in Malang, East Java. The study was conducted under the determination letter number 074/124/102.20-A/2022. To extract the active compounds from the *A. camansi* leaves, a dry powder weighing 200 grams was mixed with 96% ethanol (Merch) in a ratio of 1:10 and macerated for three cycles of 24 hours each. The resulting liquid extract was then concentrated into a thick extract using a Rotavapor® apparatus. Different concentrations of concentrated extract were prepared by weighing 2.5 grams of extract then dissolved in 500 ml, homogenized to obtain a stock solution of 5000 mg/L, then diluted to obtain The concentration of extract were 2.5 mg/L, 5 mg/L, 7.5 mg/L, and 10 mg/L.

Rotenone and *A. camansi* Treatment

To induce a Parkinson's disease model in zebrafish, a concentration of 5 μ g/L rotenone in DMSO (Sigma R8875) was added to a 2L water volume in a 25 x 16.5 x 12.5 cm aquarium, simultaneously the extract of *A. camansi* was administered in different concentrations (2.5, 5, 7.5, and 10 mg/L). The zebrafish male and female were placed in difference aquarium to compare the effects. Then, the aquarium water was refreshed every 2 days to maintain the rotenone concentration. The water temperature in the tank was tightly controlled between 24-25.5°C, and a light-dark cycle of 14:10 was established.¹⁴ Feeding the zebrafish was three times daily

with Tetra Bit and Color Tropical Flakes from Tetra Sales, Blackburn, Germany. This treatment conducted along 28 days.

Analysis of α -synuclein concentration with Immunohistochemistry (IHC) Technique

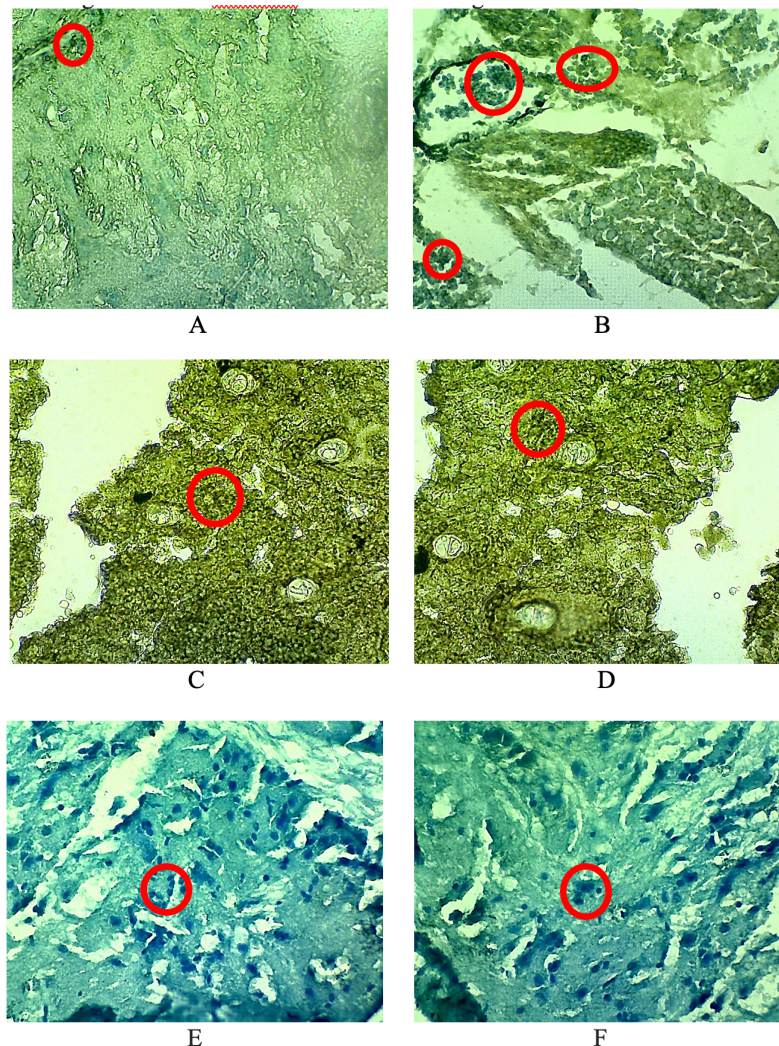
After 28 days, the animal was decapitated, the brain was isolated carefully and immediately immersed in a formalin buffer and prepared in paraffin blocks. The paraffin blocked samples were sectioned at a thickness of 0.4 mm for immunohistochemical staining. For staining, the slides were deparaffinized and subjected to the Millipore IHC procedure (Cat No. DAB 500). In this study, α -synuclein (Sigma) was used as the primary antibody, and the qualitative or quantitative expression of α -synuclein (brown color staining) was observed. Twenty fields of view in different areas were observed using a microscope on each slide at a magnification of 1000x, then quantified to obtain the average values of each treatment group were obtained from three replications (3 slides).

Data analysis

The data collected from each treatment group were subjected to statistical analysis using SPSS version 29. A one-way ANOVA test ($p < 0.05$) was employed for the analysis. The results are presented as the mean \pm standard deviation (SD) for each respective treatment group.

RESULTS AND DISCUSSION

The image below shows where the positive cells are brownish (Qualitative), then quantified to obtain the average value of α -sinuclein levels shown in Figure 2.



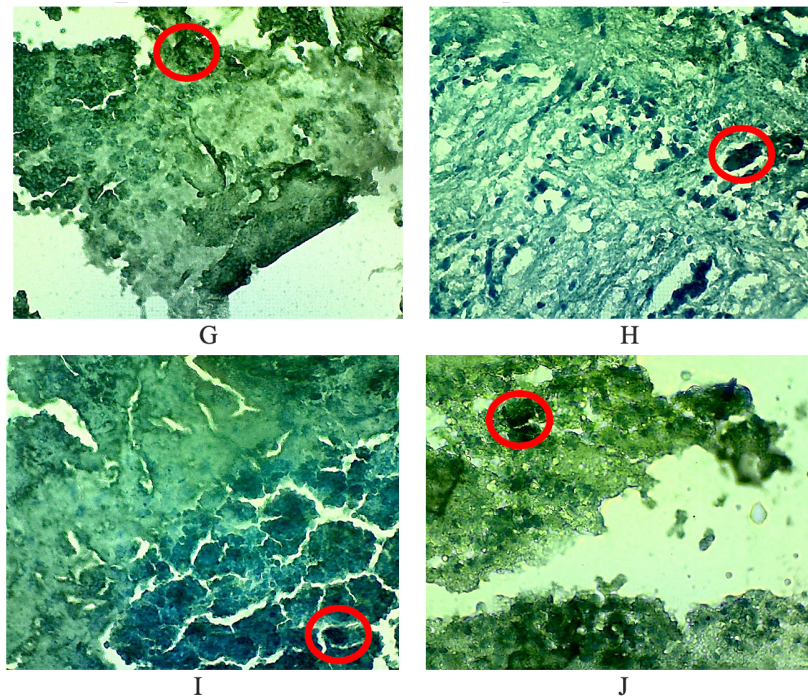


Figure 1. Staining Result of Brain Zebrafish to Evaluate Alpha-Synuclein Aggregation (brownish, in red circle). A: Control; B: Control Negative / Rotenone treatment; C: 2.5 mg/L leave extraction from adult male; D: 2.5 mg/L leave extraction from adult female; E: 5 mg/L leave extraction from adult male; F: 5 mg/L leave extraction from adult female; G: 7.5 mg/L leave extraction from adult male; H: 5 mg/L leave extraction from adult female; I. 10 mg/L leave extraction from adult male; J. 10 mg/L leave extraction from adult female

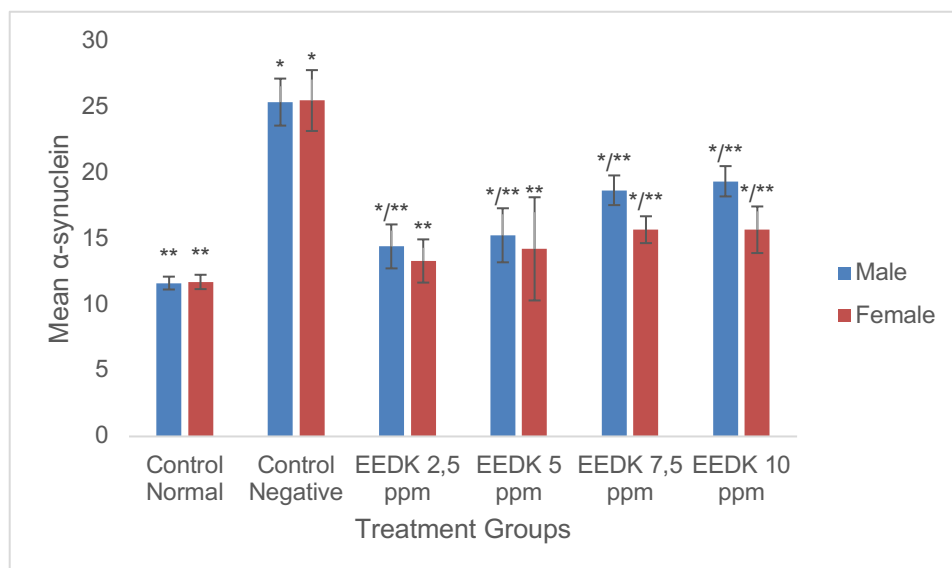


Figure 2. The Mean of α -synuclein Aggregation in Adult Zebrafish as Parkinson's Diseases Model with Ethanol Extract of *Keluwih* Leaves. *Significantly different with control normal, **Significantly different with control negative

Upon administration, rotenone notably escalated the levels of α -synuclein in both male and female adult zebrafish, presenting a significant deviation from the control group ($p < 0.05$). Contrastingly, the introduction of *A. camansi* extract resulted in a marked decline in α -synuclein levels when juxtaposed with the control group. A comparison of α -synuclein aggregation between untreated and *A. camansi*-treated samples, as visually demonstrated in Figure 1, revealed a more subdued brownish color, indicative of α -synuclein aggregation, in the latter. Statistical evaluation substantiated this visual observation, revealing a significant improvement in α -synuclein levels following *A. camansi* treatment ($p < 0.05$). These findings imply that *A. camansi* extract may have potential as a therapeutic agent in mitigating α -synuclein-related pathologies in rotenone-induced Parkinson's models in zebrafish.

Rotenone, owing to its potent lipophilic attributes, permeates cellular membranes with ease and speed, and infiltrates the brain with striking rapidity.¹⁵ The inherent toxicity of rotenone incites a domino effect of damaging consequences, foremost among them being the impairment of mitochondrial electron transport. This, in turn, results in respiratory failure and the ultimate demise of the cell, or apoptosis.¹⁶

Rotenone's initial onslaught triggers oxidative stress, culminating in alterations to DNA, lipids, and protein folding, and subsequently sparking neurodegenerative changes.¹⁷ These changes precipitate lipid modifications, thereby prompting mitochondrial dysfunction. The intricate interplay between mitochondrial damage and respiratory failure constitutes a hazardous cycle, spurred on by the dysfunction of electron transport.

Interestingly, respiration is intrinsically linked to ATP production, a cornerstone in facilitating axonal transport and cell metabolism.¹⁸ This entire gamut of deleterious conditions inexorably leads to neuronal apoptosis, especially impacting the dopaminergic neurons of the substantia nigra. This, in turn, engenders a considerable reduction in dopamine production, a process vital to the neuron's survival.¹⁹

Moreover, the synergistic action of rotenone toxicity and oxidative stress induced by mitochondrial dysfunction has the potential to modify α -synuclein formation. α -synuclein, a presynaptic protein ubiquitous in various brain regions, under physiological conditions assumes a random formation, naturally unfolding rather than aggregating.²⁰ However, upon exposure to unfavorable conditions such as low pH, organic solvents, high temperatures, metal ions, oxidative stress, and pesticides like rotenone, α -synuclein tends to aggregate in cells. This is believed to play a crucial role in the pathogenesis of Parkinson's disease (PD), as such aggregation is predominantly found in neuronal cells.²¹

Investigations into the aggregation of α -synuclein in zebrafish models have revealed that it primarily takes place in axons and is highly toxic, often preceding neuronal death.^{22,23} Such aggregation is associated with a reduced lifespan in fish models.²⁴ Further demonstrated that α -synuclein aggregation disrupts cellular microtubules and impairs mitochondrial axonal transport, owing to its high affinity binding to lipid structures such as cell membranes and organelles.²⁵ Thus, mitochondrial dysfunction in PD patients is influenced not only by rotenone exposure but also by α -synuclein aggregation.

Interestingly, the present study exhibits that the administration of an ethanol extract of *A. camansi* leaves can lower α -synuclein levels in rotenone-induced Parkinson's models in zebrafish. The improvement observed across all doses was significantly different from the control group, with the most optimal dose, based on α -synuclein measurements, being 2.5 mg/L. The constituents of the ethanol extract of *A. camansi* leaves flavonoids, alkaloids, tannins, triterpenoids, and phenolics known to be involved in the cellular pathology of Parkinson's, appear to have a synergistic effect on α -synuclein levels.

A. camansi leaves have been shown to inhibit acetylcholinesterase, the enzyme responsible for neuromelanin formation in the human brain. Neuromelanin, when increased significantly, can contribute to dopamine neurotoxicity and precipitate severe neurodegeneration. Therefore, the inhibition of neuromelanin by *A. camansi* might help stave off dopamine neurotoxicity and neurodegeneration.²⁶

Additionally, the antioxidant and anti-inflammatory properties of the *A. camansi* leaf extract could potentially stabilize the synthesis, availability, and kinetics of dopamine.¹⁸ This is further buttressed by the extract's capacity to prevent α -synuclein from aggregating, which would otherwise be neurotoxic, and inhibit the development of dopaminergic neuron degeneration.

CONCLUSION

Treatment of *keluwih* (*A. camansi*) leaves with 96% ethanol extract can be used as an alternative Parkinson's Disease ailment due to the ability of decreasing α -synuclein aggregation measurement, both male and female adult zebrafish. The lowest dose in this research of 2.5 mg/L is the optimum dose of 96% ethanol extract of *keluwih* leaves in adult male and female zebrafish. It is necessary to carry out further tests on other markers to ensure the pharmacological effects that contribute to PD pathology, and in silico tests can be carried out to determine compounds that have an effect.

AUTHORS' CONTRIBUTIONS

Marisca Evalina Gondokesumo: prepared the samples, designed the protocols, executed the protocols, wrote the manuscript, submit and revision the manuscript. Krisyanti Budipramana: reviewed and supervised the manuscript. Putu Dea Angelita Putri and Ni Putu Diah Nopitasari: data collection. Martanty Aditya and Liza Yudistira Yusan: data analytic and visualization statistically. All authors have read and approved the final manuscript.

ACKNOWLEDGEMENT

We would like to thank to Faculty of Pharmacy University of Surabaya and Faculty of Fisheries and Marine Sciences Airlangga University for facilities and material in this research.

FUNDING INFORMATION

This research received no specific grant from any funding agency in the public, commercial or not-for-profit sectors.

DATA AVAILABILITY STATEMENT

The utilized data to contribute in this research are available from the corresponding author on reasonable request.

DISCLOSURE STATEMENT

The views and opinions expressed in this article are those of the authors and do not necessarily reflect the official policy or position of any affiliated agency of the authors. The data is the result of the author's research and has never been published in other journals.

REFERENCE

1. World Health Organization. Parkinson Disease: A public health approach. *World Health Organization*. 2022;291(3):390.
2. Donadio V, Incensi A, Rizzo G, et al. The Effect of Curcumin on Idiopathic Parkinson Disease: A Clinical and Skin Biopsy Study. *J Neuropathol Exp Neurol*. 2022;81(7):545-552. doi:10.1093/jnen/nlac034

3. Robea MA, Balmus IM, Ciobica A, et al. Parkinson's disease-induced Zebrafish models: Focussing on oxidative stress implications and sleep processes. *Oxid Med Cell Longev*. 2020;2020:1-15. doi:10.1155/2020/1370837
4. Razali K, Othman N, Mohd Nasir MH, et al. The promise of the zebrafish model for Parkinson's disease: Today's science and tomorrow's treatment. *Front Genet*. 2021;12. doi:10.3389/fgene.2021.655550
5. Binienda ZK, Sarkar S, Mohammed-Saeed L, et al. Chronic exposure to rotenone, a dopaminergic toxin, results in peripheral neuropathy associated with dopaminergic damage. *Neurosci Lett*. 2013;541:233-237. doi:10.1016/J.NEULET.2013.02.047
6. Ma'arif B, Maimunah S, Muslikh FA, et al. Efek ekstrak daun Marsilea crenata Presl. pada aktivitas lokomotor ikan zebra. *FARMASIS: Jurnal Sains Farmasi*. 2022;3(1):18-24. doi:10.36456/FARMASIS.V3I1.5389
7. Oliveira RF. Mind the fish: zebrafish as a model in cognitive social neuroscience. *Front Neural Circuits*. 2013;7:1-15. doi:10.3389/fncir.2013.00131
8. Martinez TN, Greenamyre JT. Toxin models of mitochondrial dysfunction in Parkinson's disease. *Antioxid Redox Signal*. 2012;16(9):920-934. doi:10.1089/ars.2011.4033
9. Prakash O, Kumar A, Gupta R. Evaluation of anticonvulsant activity of Artocarpus heterophyllus Lam. leaves (Jackfruit) in mice. *Scholars Research Library*. 2013;5(1):217-220. Accessed August 26, 2023. www.scholarsresearchlibrary.com
10. Jagtap UB, Bapat VA. Artocarpus: A review of its traditional uses, phytochemistry and pharmacology. *J Ethnopharmacol*. 2010;129(2):142-166. doi:10.1016/j.jep.2010.03.031
11. Solichah AI, Anwar K, Rohman A, Fakhrudin N. Profil fitokimia dan aktivitas antioksidan beberapa tumbuhan genus Artocarpus di Indonesia. *JFood PharmSci*. 2021;9(2):443-460. www.journal.ugm.ac.id/v3/JFPA
12. Das S, Laskar MA, Sarker SD, et al. Prediction of anti-Alzheimer's activity of flavonoids targeting acetylcholinesterase in silico. *Phytochemical Analysis*. 2017;28(4):324-331. doi:10.1002/pca.2679
13. Outeiro TF, Ferreira JJ. Zebrafish as an Animal Model for Drug Discovery in Parkinson ' s Disease and Other Movement Disorders : A Systematic Review. 2018;9(June). doi:10.3389/fneur.2018.00347
14. Khotimah H, Ali M, Sumitro SB, Widodo MA. Decreasing α -synuclein aggregation by methanolic extract of Centella asiatica in zebrafish Parkinson's model. *Asian Pac J Trop Biomed*. 2015;5(11):948-954. doi:10.1016/j.apjtb.2015.07.024
15. Irwin MH, Parameshwaran K, Pinkert CA. Mouse models of mitochondrial complex I dysfunction. *Int J Biochem Cell Biol*. 2013;45(1):34-40. doi:10.1016/j.biocel.2012.08.009
16. Ott K. Rotenone. A brief review of its chemistry, environmental fate, and the toxicity of rotenone formulations. *Environmental Science*. Published online 2006.
17. Fleisch VC, Fraser B, Allison WT. Investigating regeneration and functional integration of CNS neurons: Lessons from zebrafish genetics and other fish species. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. 2011;1812(3):364-380. doi:10.1016/j.bbadis.2010.10.012
18. Khotimah H, Sumitro SB, Aris Widodo M. *Zebrafish Parkinson's Model: Rotenone Decrease Motility, Dopamine, and Increase α -Synuclein Aggregation and Apoptosis of Zebrafish Brain*. Vol 8.; 2015.
19. Sherer TB, Betarbet R, Testa CM, et al. Mechanism of toxicity in rotenone models of Parkinson's disease. *The Journal of Neuroscience*. 2003;23(34):10756-10764. doi:10.1523/JNEUROSCI.23-34-10756.2003

20. Uversky VN, Eliezer D. *Biophysics of Parkinson's Disease: Structure and Aggregation of α -Synuclein*. Vol 10.; 2009.
21. He Q, Song N, Xu H, Wang R, Xie J, Jiang H. Alpha-synuclein aggregation is involved in the toxicity induced by ferric iron to SK-N-SH neuroblastoma cells. *J Neural Transm (Vienna)*. 2011;118(3):397-406. doi:10.1007/s00702-010-0453-0
22. Lopez A, Lee SE, Wojta K, et al. A152T tau allele causes neurodegeneration that can be ameliorated in a zebrafish model by autophagy induction. *Brain*. 2017;140(4):1128-1146. doi:10.1093/brain/awx005
23. Weston LJ, Cook ZT, Stackhouse TL, et al. In vivo aggregation of presynaptic alpha-synuclein is not influenced by its phosphorylation at serine-129. *Neurobiol Dis*. 2021;152. doi:10.1016/j.nbd.2021.105291
24. Prabhudesai S, Sinha S, Attar A, et al. A novel "Molecular Tweezer" inhibitor of α -Synuclein neurotoxicity in vitro and in vivo. *Neurotherapeutics*. 2012;9(2):464-476. doi:10.1007/s13311-012-0105-1
25. Mahul-Mellier AL, Burtscher J, Maharjan N, et al. The process of Lewy body formation, rather than simply α -synuclein fibrillization, is one of the major drivers of neurodegeneration. In: *The Proceedings of the National Academy of Sciences (PNAS)*. Vol 117. ; 2020:4971-4982. doi:10.6084/m9.figshare.11842389.v2
26. Orhan IE. Centella asiatica (L.) urban: From traditional medicine to modern medicine with neuroprotective potential. *Evidence-Based Complementary and Alternative Medicine*. 2012;2012:1-8. doi:10.1155/2012/946259